

Analysis of flavonoids in grape leaves by HPLC-DAD-MS/MS

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Introduction

Flavonoids are water soluble polyphenolic molecules which belong to a group of plant metabolites thought to provide health benefits through cell signalling pathways and antioxidant effects. Flavonoids are the most abundant and widely studied, and have enjoyed greater attention among grape researchers¹⁾.

Experimental methods

Grape leaves of various grape varieties obtained from the Research Institute of Viticulture and Enology in Slovakia were analysed. Samples were collected in September 2013. Leaves were dried and frozen (–20 °C) until treated prior to analysis. Samples were homogenized and 2.5 g of each sample was extracted with a 40 ml mixture of methanol/water 80 : 20 (v/v) for 120min and 2 × 20 ml dichloromethane was used for sample clean-up. 2.5 ml of the water-methanolic part was dried and dissolved in the mobile phase. Analysis was performed on an LC Agilent 1200 Infinity System (Agilent Technologies, USA) with an Agilent 1260 Infinity Diode Array Detector and an Agilent 6520 Accurate-Mass Quadrupole Time-of-Flight. Separation column was Kromasil C18 (4.0 × 150 mm, 3.5 μm). Mobile phase consisted of acetonitrile and formate buffer (ammonium formate of the concentration 10 mmol/l and pH adjusted to 3.1 by formic acid). Gradient elution 10–95% of acetonitrile in 62 min was used.

MS Detection conditions were: negative mode (ESI–), capillary voltage: 3.5 kV, drying gas temperature: 300 °C, drying gas flow rate: 10 l/min, nebulizer pressure: 40 psi, fragmentor voltage: 140 V, collision energy: 20 eV.

Results and discussion

Identification of the separated compounds was carried out using a UV, MS and MS/MS spectra. 7 quercetin and 4 kaempferol glycoside flavonoids (quercetin-3-*O*-rutinoside, quercetin-glucuronide, quercetin-3-*O*-galactoside, quercetin-3-*O*-glucoside, quercetin-pentoside 1, kaempferol-3-*O*-rutinoside, kaempferol-hexoside 1, quercetin-

-pentoside 2, quercetin-3-*O*-rhamnoside, kaempferol-hexoside 2, kaempferol-deoxyhexoside) were identified. A DAD detector and external calibration curves of quercetin and kaempferol (in the range 5–100 μg/ml) were used for quantification. 9 of the 13 identified compounds could be quantified (Table 1).

Table 1. Content of glycoside flavonoids in grape leaves:

Compound	Range of content (μg/g)
quercetin-3- <i>O</i> -rutinoside	41.79–1576.58
quercetin-glucuronide	1385.49–7284.86
quercetin-3- <i>O</i> -galactoside	95.49–1152.52
quercetin-3- <i>O</i> -glucoside	83.51–4665.55
quercetin-pentoside 1	< LOD–1022.68
kaempferol-3- <i>O</i> -rutinoside	< LOQ–913.51
kaempferol-hexoside 1	< LOQ–1348.04
quercetin-3- <i>O</i> -rhamnoside	< LOQ–1307.75
kaempferol-hexoside 2	< LOQ–2380.98

It was found that the highest contained flavonoid was quercetin-glucuronide, the content of which ranged from 1385 to 7284 μg/g. The amount of other components varied from 41.79 μg/g to the maximum value of 4665 μg/g. The method validation confirmed sufficient linearity, precision, accuracy and separation efficiency. Low limits of detection and limit of quantification (LOD = 1.12 μg/g, LOQ = 3.73 μg/g for quercetin and LOD = 2.35 μg/g, LOQ = 7.84 μg/g for kaempferol) were detected.

Conclusions

It was confirmed that the proposed method is suitable for a comprehensive analysis of complex plant matrices, such as grape leaves. The results of research will contribute to the creation of profiles of pharmaceutically active substances contained in the leaves of nearly 30 Slovak vine varieties that could be used in the future to prepare nutraceuticals.

Research was supported by grant APVV-0550-11.

Conflicts of interest: none.

References

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