Estimation of lipohydrophilic properties of molecules with potential β₃-agonistic activity

ĽUBICA HAVRANOVÁ SICHROVSKÁ[1], LUKÁŠ STANZEL[1], IVAN MALÍK[1], MATEJ MARUNIAK[1], IVA KAPUSTÍKOVÁ[1], EVA SEDLÁROVÁ[1], JOZEF CSÖLLEI[2]

[1]Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Comenius University, Slovak Republic
[2]Department of Chemical Drugs, Faculty of Pharmacy, University of Veterinary and Pharmaceutical Sciences, Brno, Czech Republic

Introduction

Lipophilicity is considered to be a very important molecular descriptor. It plays a crucial role in determining pharmacokinetic ADMET-properties (absorption, distribution, metabolism, excretion, and toxicity) and the pharmacodynamic profile, which correlates well with their bioactivity in the end[1]. Successful drug development requires efficient delivery to target sites as the drug must be able to reach a specific biophase by crossing several biomembranes by passive and/or active transport. A predominant factor influencing the pharmacokinetic behaviour is lipophilicity. For example, commonly used β-blockers, which are structurally similar to β₃-adreneregic receptor agonists, may be divided into lipophilic and hydrophilic drugs, or are in an intermediate position[2]: sotalol (log P_{oct} = –0.79), atenolol (log P_{oct} = 0.23), nadolol (log P_{oct} = 0.71), practolol (log P_{oct} = 0.76), pindolol (log P_{oct} = 1.75), acebutolol (log P_{oct} = 1.87), timolol (log P_{oct} = 2.10), metoprolol (log P_{oct} = 2.15), alprenolol (log P_{oct} = 2.61) and propranolol (log P_{oct} = 3.65). Lipophilicity of compounds can influence oral absorption, diffusion through biolog ical barriers (e.g. placenta or blood/brain), degree of metabolism/renal elimination, plasma half-life, receptor selectivity, or tissue concentration[3]. Highly lipophilic drugs are insoluble in aqueous media and bind strongly to plasma proteins, which results in a low free blood concentration, and are distributed only into lipid bilayers. On the other hand, highly polar compounds cannot be absorbed through the gut wall because of lower membrane solubility. Keeping the optimal lipophilicity range can lead to an improvement of therapeutic efficacy and side-effect profiles of new drugs[4].

Experimental methods

To describe the transfer of a substance from the aquatic environment into an organism and its bioaccumulation potential, the partition coefficient of a substance between water and a lipophilic solvent (octan-1-ol, cyclohexane, heptane) was determined.

PharmDr. Ľubica Havranová Sichrovská (✉)
Katedra farmaceutickej chémie
Odbojárov 10, 832 32 Bratislava, Slovak Republic
e-mail: l.sichrovska@gmail.com

Partition coefficient (P) is defined as the ratio of the equilibrium concentrations (C_i) of a dissolved substance in a two-phase system consisting of two immiscible solvents[5]:

$$P = C_{LS}/C_W, \qquad [1]$$

where C_{LS} is the concentration of a compound in the lipophilic phase and C_W is the concentration of a compound in the aqueous phase. Partition coefficient is usually given in the form of its log arithm to the base ten (log P).

Studied compounds

Chemical structure and the basic physicochemical parameters of the studied substances *BL-14S2-BL-44S2* (chemically 3-{4-[(alkoxycarbonyl)amino]phenoxy}-*N*--{2-[4-(aminosulfonyl)phenyl]ethyl}-2-hydroxypropan--1-ammonium chlorides) are shown in Table 1.

Devices

An analytical balance Chyo JL-180 (Chyo Balance Corporation, Japan), a mechanical shaker, a UV spectrophotometer (Shimadzu, UV-1800, Japan), a pH-meter (Hanna Instruments, Slovak Republic).

Chemicals

The aqueous phase was represented in all cases by phosphate buffer prepared from a water solution of disodium hydrogen phosphate, p.a. (CentralChem, Slovak Republic) with the concentration c = 0.2 mol · l⁻¹ and a water solution of citric acid, p.a. with the concentration c = 0.1 mol · l⁻¹. Measurements were performed under equilibrium conditions at pH = 7.4. The lipophilic phase was represented by high purity analytical grade octan-1-ol (Merck, Germany), cyclohexane (CentralChem, Slovak Republic) and heptane (CentralChem, Slovak Republic).

Estimation of partition coefficient log P2

In the present study, the generally accepted and well-known shake-flask method for obtaining the log P values in three mediums was used. The first one was octan-1-ol/phosphate buffer (log P_O), second cyclo-hexane/phosphate buffer (log P_C) and the third was heptane/phosphate buffer (log P_H). The authors prepared 50 ml of basic solutions of the studied compounds *BL-14S2-BL-44S2* in phosphate buffer at pH = 7.4 with the concentration c = 5.10⁻⁵ mol · l⁻¹ and measured their absorbance A_1. To 10 ml of the solution, 0.5 ml of the lipophilic medium represented by octan-1-ol, cyclohexane and heptane, respectively, was added. This system was

shaken for 1h, the phases of the solvent system were mutually saturated and after that settled for 1 h, then the lipophilic and aqueous phases were separated. The absorbance A_2 of the aqueous phase was measured. The values of absorbance were in both cases measured at the wavelength λ = 227 nm. The data of log P_O, log P_C, log P_H for each compound were calculated from the equations 2–4[6)]:

$$P_{exp} = (1000 \; g) - (a \times c_{H_2O} \times M_r) / b \times c_{H_2O} \times M_r \quad [2]$$

$$c_{H_2O} = A2 \quad\quad\quad\quad\quad\quad\quad\quad\quad\quad\quad [3]$$

$$ε = A_1 /c, \quad\quad\quad\quad\quad\quad\quad\quad\quad\quad [4]$$

where g is the weight of the studied compound in grams in 10 ml of measured solution, a is the number of millilitres of the aqueous phase, b is the number of millilitres of the lipophilic phase, M_r is the relative molecular weight of the compound under study, c_{H2O} is the amount of the inspected compound in the aqueous phase after shaking, c is the concentration of the measured solution expressed in mol · l⁻¹.

Results and discussion

This short research paper was focused on the estimation of lipophilicity of four newly synthesized substances potentially active as β₃-adrenergic receptor agonists. Solubility in lipids refers to the ability of a compound to dissolve in fats, oils, lipids and non-polar solvents[7)]. The experimentally estimated partition coefficient (log P) is routinely used as an assessment of lipid solubility *in vivo* and it is a key event of molecular desolvation in the transfer from aqueous phases to the cell membrane and protein bindings. The values of log P were experimentally determined in three systems, whereby the aqueous phase was always composed of phosphate buffer with pH = 7,4 and the lipophilic phase was formed of octan-1-ol, cyclohexane and heptane. Octan-1-ol represents a simple model of the cell phospholipidic membrane. The log P data acquired from the systems with cyclohexane and heptane signify the margin of penetration through the *stratum corneum* and blood-brain barrier. Octan-1-ol has eight atoms of carbon which are responsible for its lipophilic properties and on the other hand, it has also hydroxyl functionality which is accountable for its hydrophilic attribute. Cyclohexane is a lipophilic solvent composed of a planar six-membered cycle; therefore the molecules with benzene rings should easily incorporate into this system. Heptane is also a lipophilic solvent but it is composed of linear aliphatic chains. Compounds with aromatic scaffold cannot incorporate into heptane molecules so easily[8)].

The evaluated compounds *BL-14S2-BL-44S2* are structurally based on the aryloxyaminopropanol pharmacophore bearing a benzene sulfonamide fragment in the basic part of the molecule differing from each other in the alkoxylcarbonylamino moiety. The presence of two aromatic rings is responsible for their lipophilic character. Furthermore, with elongation of the alkoxycarbonylamino fragment attached to the aromate, lipophilicity enhances. On the other hand, the presence of quaternary ammonium,

Table 1. General characterization of investigated molecules BL-14S2-BL-A4S2

	R	Formula	M_r	m.p. (°C)
BL-14S2	CH_3	$C_{19}H_{26}ClN_3O_6S$	459.944	219–222
BL-24S2	C_2H_5	$C_{20}H_{28}ClN_3O_6S$	473.971	210–214
BL-34S2	C_3H_7	$C_{21}H_{30}ClN_3O_6S$	487.997	220–223
BL-44S2	C_4H_9	$C_{22}H_{32}ClN_3O_6S$	502.024	222–224

Table 2. Values of absorbances for calculation log P_O, log P_C, log P_H for studied compounds BL-14S2--BL-44S2

Entry	A_1	A_{2O}	A_{2C}	A_{2H}
BL-14S2	1.130	0.803	1.128	1.129
BL-24S2	1.097	0.516	1.096	1.096
BL-34S2	1.117	0.706	1.116	1.118
BL-44S2	0.869	0.253	0.806	0.812

A_1 – absorbance measured before shaking, A_{2O} – absorbance measured after shaking for the system octan-1--ol/phosphate buffer, A_{2C} – absorbance measured after shaking for the system cyclohexane/phosphate buffer, A_{2H} – absorbance measured after shaking for the system heptane/phosphate buffer

Table 3. Experimentally estimated values of partition coefficients for inspected compounds BL-14S2-BL-A4S2 in octan-1-ol / phospate buffer (log P_O), cyclohexane / phosphate buffer (log P_C) and heptane / phosphate buffer (log P_H)

Entry	log P_O	log P_C	log P_H
BL-14S2	0.90	–	–
BL-24S2	1.07	–	–
BL-34S2	1.35	–	–
BL-44S2	1.70	0.22	0.17

hydroxyl and sulfonamide groups is responsible for their hydrophilicity. Whereas, cyclohexane and heptane are highly lipophilic solvents, only the substance *BL-44S2* with a butoxylcarbonylamino moiety was able to penetrate into them. The partition coefficient in the medium cyclohexane/phosphate buffer (log P_C) system was of the value of 0.22 and was higher than the partition coefficient estimated in heptane/phosphate buffer, which was of the value of 0.17, in consequence of a different structural character of the molecules of the solvents mentioned above. Into the non-polar environment of octan-1-ol all of the substances were able to penetrate and the values of log P_O ranged from 0.90 to 1.70 (Table 3), whereby with an elongation of the alkoxylcarbonylamino fragment, the log P_O values increased constantly.

Conclusions

In conclusion, following the obtained results it can be assumed that the currently investigated compounds *BL-14S2-BL-44S2* which could be classified as potential β_3-adrenergic agonists, are not highly lipophilic and according to that, they are unlikely to cross the blood-brain barrier and cause some serious undesirable side effects on the CNS. It could be hypothesized that they would not be extensively metabolized, have not poor absorption, solubility, low bioavailability and a shorter half-life compared to their lipophilic analogues.

Conflicts of interest: none.

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