

Thin-layer chromatography application for the standardization of *Sophora* flowers

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Introduction

Herbal products contain multiple constituents that might be responsible for its therapeutic effects. Thus it is necessary to define as many constituents as possible in order to understand and explain their bioactivity. According to this concept, a chemical profile for herbal product is constructed and compared with the profile of a clinically proven reference product. Since many of these preparations contain flavonoids, it is essential to have adequate analytical techniques at hand for this class of natural products. Various analytical methods exist for flavonoids definition.

Paper chromatography is widely used for the analysis of flavonoids, but now there are more modern methods of analysis, and simple and inexpensive thin-layer chromatography (TLC) is one of them.

There are many kinds of the systems solvent/eluent used for the separation of flavonoids using TLC. With regard to detection, flavonoids spot on TLC plates produce a yellow-brown spots white background when reacted with iodine vapour. Flavonoids may appear as dark spots on a green background fluoresce when observed in UV light at 254 nm UV-plates containing a fluorescent indicator (such as silica gel F254). If under 365 nm UV light, spot colours depending on the structure of flavonoids can be yellow, green or blue fluorescent. It would be more clear and intense after being sprayed with the reagent.

The aim of our work was to develop an identification method of flavonoids in *Sophora japonica* flowers, presented in Ukraine for the creation of national regulatory documents. For the basis we took the method of flavonoids determination displayed in the European Pharmacopoeia (EP) 8.3 Monograph “*Sophora* Flower”.

Experimental methods

For the preparation of the test solutions to 1 g of the powdered herbal drug (355) (2.9.12) we added 5.0 mL of methanol R, sonicated for 10 min and then filtered.

For the preparation of the reference solutions, standard samples of 10 mg hyperoside R and 10 mg rutin R which was dissolved in 10 mL methanol R were used.

The solutions were applied as bands of 10 µL on the starting line of the chromatographic silica gel plate R

(5–40 µm). Then the plate was immersed in the solvent system: anhydrous formic acid R, water R, ethyl acetate R (10 : 10 : 80). Development over the path of 10 cm. After that, the plate was dried in air.

Detection was performed using a 10 g/L solution of diphenylboric acid aminoethyl ester R in methanol R and then with a 50 g/L solution of macrogol 400 R in methanol R. Next, the plate was dried in air for 30 minutes and examined in ultraviolet light at 365 nm. The results were evaluated by viewing the plate under UV light at 365 nm.

Results and discussion

For the analysis of the obtained results, the size, colour, fluorescence zones, location and clarity of their separation on the chromatogram of the reference solution and the test solution were compared (Fig. 1).

Top of plate	
	an orange-yellow zone
	a brown zone
Hyperoside: a yellowish-orange zone	2 green zones
Rutin: an orange-yellow zone	A very intense orange-yellow zone (rutin)

Fig. 1. The scheme chromatogram of flavonoids in the flowers of *Sophora japonica*

In the resulting chromatogram, a zone of orange-yellow at the zone level of the standard sample rutin that matched its size and intensity of colour was observed. Above the rutin zone there were two areas of green colour that may

fit kaempferol, isorhamnetin. Approximately at the zone level of hyperoside there was a brown zone. Closer to the finish line we observed an area of orange-yellow colour which may be due to the presence of quercetin in *Sophora japonica* flowers.

Performed chromatographic analysis was characterized by a high quality of the obtained chromatograms, a clear distribution of substances and reproducibility.

Conclusions

Thus, in the flowers of *Sophora japonica* flavonoid substances of nature with reliable determinations of the contents of rutin and hyperoside were identified. Determination was performed using standard samples.

This TLC method, which is presented in the EPh 8.3 Monograph “*Sophora Flower*”, can be proposed for the introduction in the domestic regulatory documentation in the form of monographs of the State Pharmacopoeia of Ukraine “*Sophora Flower*”. Conditions of the chromatographic analysis are optimal for their application in the analysis of *Sophora japonica*, represented in Ukraine.

In favour of this technique it should also be noted that, in the development of monographs of the State Pharmacopoeia of Ukraine on medicinal herbs, it primarily deals with the qualitative and quantitative analysis techniques presented in the EPh, due to the harmonization of the State Pharmacopoeia of Ukraine with the EPh and the desire to fit modern quality requirements pertaining to medicinal plants.

Conflict of interest: none.

Reference

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